

## Application note No. 0260. Rev. 1.1

**NucleoCounter® NC-250™****Viability and Cell Count of Aggregated Mammalian Cells using NC-Slide A2™****Product description**

The NucleoCounter® NC-250™ system enables the user to perform automated cell counting and analyses of a broad range of mammalian cells.

**Application**

The NC-Slide A2™, **Solution 10** and **Solution 12** used together with the NucleoCounter® NC-250™ facilitates determination of the viability and cell concentration of aggregating cells. Treatment of cells with **Solution 10** facilitates disaggregation of cell aggregates resulting in single cell suspensions. At the same time **Solution 10** also enables staining of all cells with DAPI. One sample is analyzed in approximately 90 seconds.

**Introduction**

In order to determine the total cell concentration, a sample containing cells in suspension is mixed with **Solution 10** (lysis buffer) and **Solution 12** (staining solution) and loaded into a NC-Slide A2™. **Solution 12** contains DAPI, which stains all cell nuclei after lysis with **Solution 10**. When the cell sample is not treated with **Solution 10** the dye only stains the non-viable cells. After loading the NC-Slide A2™ it is placed in the NucleoCounter® NC-250™ where viability and cell concentration is determined.

The nominal depth of the chambers in the NC-Slide A2™ is 100 µm, with 90 % of all chambers being in the range from 90-110 µm.

**Procedures**

If the cell line to be investigated is adherent or semi-adherent, then start by getting all cells into suspension using the preferred method of your laboratory (e.g. trypsin/EDTA treatment).

**Materials needed**

- Cells to be counted
  - NC-Slide A2™
  - **Solution 10**
  - **Solution 12**
1. Two cell samples is prepared – one is treated with both **Solution 12** and **Solution 10** to determine the total cell concentration – the other cell sample is only treated with **Solution 12** to determine the concentration of the non viable cells.
  2. Sample 1: Total cell count
    - a. Add one volume of **Solution 12** into 99 volumes of **Solution 10**. E.g., per cell sample add 10 µl of **Solution 12** to 990 µl **Solution 10**.
    - b. The original cell suspension is mixed to obtain a homogenous suspension. Pipette a representative cell sample from the cell suspension into a microcentrifuge tube (e.g. 100 µl).
    - c. Add one volume of the mixture of **Solution 12** and **Solution 10** to the microcentrifuge tube with the cell sample. E.g., if the volume of the cell sample is 100 µl then add 100 µl of the mixture of **Solution 12** and **Solution 10**. Mix by pipetting.
    - d. Load ~30 µl of the cell sample for the total cell count into chamber 1 of the slide.
  3. Sample 2: Non viable cell count:
    - a. The original cell suspension is mixed again to obtain a homogenous suspension. Pipette a representative cell sample from the cell suspension into a microcentrifuge tube (e.g. 199 µl).

- b. Add one volume of **Solution 12** to 199 volumes of cells sample in the microcentrifuge tube. E.g., if the volume of the cell sample is 199  $\mu$ l then add 1  $\mu$ l of **Solution 12**. Mix by pipetting.
  - c. Load  $\sim$ 30  $\mu$ l of the sample for the non-viable cell count into chamber 2 of the slide.
4. Place the loaded slide on the tray of the NucleoCounter<sup>®</sup> NC-250™ and select “**Viability and Cell Count - Aggregated Cells Assay**” and press RUN.

After approximately 90 seconds the cell concentrations (cells/ml) of the total cell count and the viability are displayed. The displayed total cell concentration has been compensated for the dilution caused by addition of **Solution 10** and **Solution 12**. Analogous, the cell concentration for the non-viable cells has been compensated for the dilution caused by the addition of **Solution 12**. If the cell sample has been further diluted or concentrated and the user has entered the volumes into the user interface the dilution factor has also been taken into account and the returned cell concentration is for the original cell concentration.

### Notes

To assure reliable results, it is recommended that the cell concentration of the counted cell suspension should be in the range of  $5 \cdot 10^4$  cells/ml to  $5 \cdot 10^6$  cells/ml. If the cell concentration is above  $5 \cdot 10^6$  cells/ml, the cell suspension can be diluted with growth media or PBS to achieve the desired concentration. The diluted cell sample is then treated as described above. By inserting the value for the dilution volume in the dilution field on the user interface the returned cell concentration is for the original cell sample.

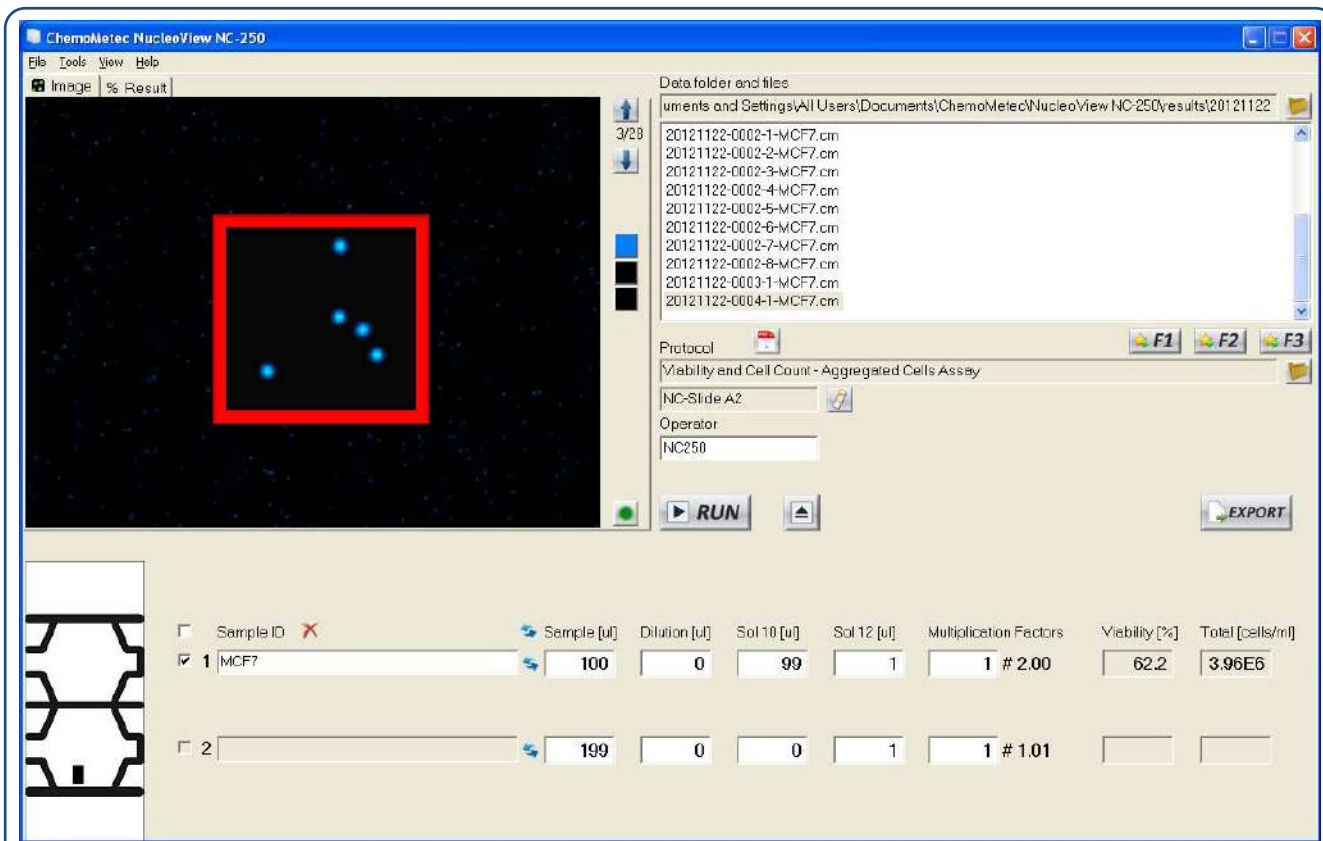
If the concentration of cells is below  $5 \cdot 10^4$  cells/ml then the cell concentration may be increased by centrifugation followed by resuspension of the pellet using growth media or PBS. The resuspended cell sample is then treated as described in the procedure. By inserting a negative value representing the volume removed from the sample in the dilution field on the user interface the returned cell concentration is for the original cell sample.

### Viability

The viability is calculated as follows:

$$\% \text{ viability} = \frac{C_t - C_{nv}}{C_t} \cdot 100\%$$

<b>% viability</b>	The percentage of viable cells in the cell suspension
<b>C<sub>t</sub></b>	The total concentration of cells
<b>C<sub>nv</sub></b>	The concentration of non-viable cells



Determination of viability and cell concentration of aggregated MCF7 cells. The cells for the total count were disaggregated and stained by adding a mixture of **Solution 10** and **Solution 12**. The cells for the dead count were only treated with **Solution 12**. The cell populations are stained with DAPI and appear blue. An insert shows a close up of parts of the images. The results are presented at the bottom right and extended results are presented in the result tab page.

## Troubleshooting

### Inaccurate and imprecise counting:

When setting up a new cell line it is important to inspect that the cell line is counted correctly. The cells included in the total count can be marked by clicking on the overlay button in the bottom right corner of the image. Visual inspect the image to evaluate in the vast majority of the cells has been counted correctly. If this is not the case right click on the image file in question and choose "Show Data in Plot Manager". Inspect the gates displayed in the Plot Manager. If the gating is inappropriate right click on the image file in question again and choose "Start Protocol Adaptation Wizard". Adapt the gate(s) to cover the cell population (do not include debris and very large objects) and save the changes to a new protocol. Note that the user is responsible for defining appropriate gating of the particular cell line.

### Inappropriate loading of the NC-Slides:

Due to variations in chamber volumes the exact amount needed to fill the chamber may vary. Make sure that the chamber is completely filled and that no excess liquid spreads into other chambers or onto the top of the coverslip. Furthermore, avoid introduction of air bubbles into the chambers. Insufficient filling and air bubbles may cause cell movement compromising the quality of the image analysis.

Warning that the cell concentration of non viable cells is higher than the total cell concentration:

Make sure the problem is not due to interchanged samples of the total count sample treated with a mixture of **Solution 10** and **Solution 12** and the non-viable count sample only treated with **Solution 12**. If the samples have not been interchanged the continued warning can be due to a very high frequency of non-viable cells in the sample.

Make sure the problem is not due to wrong entered volumes on the user interface.

**Handling and storage**

For handling and storage of ChemoMetec instruments and reagents, cassettes and NC-slides refer to the corresponding product documentation. For other reagents refer to the material data sheet from the manufacturer of the reagents and chemicals.

**Warnings and precautions**

For safe handling and disposal of the ChemoMetec reagents, cassettes and NC-slides refer to the corresponding product documentation and the NucleoCounter® NC-250™ user's guide. For other reagents refer to the safety data sheet from the manufacturer of the reagents and chemicals required for this protocol. Wear suitable eye protection and protective clothes and gloves when handling biologically active materials.

**Limitations**

The NucleoCounter® NC-250™ system is FOR RESEARCH USE ONLY. NOT FOR DIAGNOSTIC OR THERAPEUTIC USE. The results presented by the NucleoCounter® NC-250™ system depend on correct use of the reagents, NC-slide and the NucleoCounter® NC-250™ instrument and might depend on the type of cells being analyzed. Refer to the NucleoCounter® NC-250™ user's guide for instructions and limitations.

**Liability disclaimer**

This application note is for RESEARCH PURPOSES ONLY. It is not intended for food, drug, household, or cosmetic use. Its use must be supervised by a technically qualified individual experienced in handling potentially hazardous chemicals. The above information is correct to the best of our knowledge. Users should make independent decisions regarding completeness of the information based on all sources available. ChemoMetec A/S shall not be held liable for any damage resulting from handling or contact with the above product.

**Product disclaimer**

ChemoMetec A/S reserves the right to introduce changes in the product to incorporate new technology. This application note is subject to change without notice.

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